

1 **Doberman pinschers present autoimmunity associated with functional autoantibodies:**
2 **a model to study the autoimmune background of human dilated cardiomyopathy**

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13 **Keywords:** anti-beta1-adrenergic receptor autoantibodies, Doberman cardiomyopathy,
14 functional autoantibodies, human dilated cardiomyopathy, anti-muscarinic receptor 2
15 autoantibodies

16

17 **Short title:** Doberman cardiomyopathy for modeling human immune-mediated dilated
18 cardiomyopathy

19

20 **Potential conflict of interest:**

21 The authors declare that the study was not funded by any public, commercial or private funds.
22 Gerd Wallukat, Niels-Peter Becker, Katrin Wenzel, Johannes Müller and Ingolf Schimke as
23 employees and shareholder (GW, JM, IS) of Berlin Cures GmbH declare that Berlin Cures
24 GmbH only provided support in the form of salaries and research materials but did not have
25 any additional role in the study design, data collection, analysis and statistical evaluation,
26 decision to publish or preparation of the manuscript. Anna Fritscher and Gerhard Wess declare
27 that they have no conflict of interest.

28

29 Berlin Cures GmbH, a spin-off company, founded in September 2014 for the commercial exploitation of
30 patents held by Charité – Universitätsmedizin Berlin and Max-Delbrück-Center Berlin, Germany.

31 **Abstract**

32 **Background**

33 Autoimmunity associated with autoantibodies directed against the β 1-adrenergic receptor (β 1-
34 AAB) is increasingly accepted as driving human dilated cardiomyopathy (DCM). Unfortunately,
35 animal models of DCM are lacking, preventing our knowledge about β 1-AAB autoimmunity in
36 DCM from being extended and hindering the development of related treatment strategies.

37 **Objectives**

38 To introduce an animal model, we studied Doberman pinschers, which develop
39 cardiomyopathy (DoCM), with similarities to human DCM, with regard to their β 1-AAB
40 autoimmunity.

41 **Methods**

42 Eighty-seven DP with DoCM and 31 (at enrolment) healthy controls were analyzed for β 1-AAB;
43 the receptor binding site and sensitivity to inhibition were determined. In controls who
44 developed cardiomyopathy during the follow-up, β 1-AAB were analyzed during the DoCM
45 progress.

46 **Results**

47 Fifty-nine (67.8%) DoCM dogs and 19 (61.3%) controls were β 1-AAB positive. Excluding the
48 9 controls who developed DoCM in the follow-up, β 1-AAB positivity tended to be more
49 pronounced in DoCM.

50 From the controls who developed DoCM, 8 were β 1-AAB positive ($p=0.044$ vs. dogs remaining
51 healthy); their β 1-AAB level increased with the cardiomyopathy progress. Overall mortality and
52 mortality exclusively due to cardiac reasons during the study period, were higher ($p=0.002$;
53 $p=0.0037$) in β 1-AAB positive dogs. The dogs' β 1-AAB targeted a specific epitope centralized
54 on the second extracellular receptor and were sensitive to inhibition by drugs already
55 successful tested for the corresponding human autoantibody.

56 **Conclusions**

57 Doberman pinschers presented β 1-AAB associated autoimmunity similar to that driving the
58 pathogenesis of human DCM. Consequently, DP could remove the lack of animal models
59 available for studying β 1-AAB autoimmunity in DCM.

60 **Introduction**

61 Autoimmunity is increasingly accepted as the origin or amplifier of heart failure (1). For
62 cardiomyopathies, preferentially for dilated cardiomyopathy (DCM) with idiopathic nature (as
63 recently for the US re-calculated prevalence of 1 in 250-400 individuals (2)) and that
64 secondarily to non-ischemic causes, e.g. infectious diseases such as myocarditis, several
65 autoantibodies directed against cardiac antigens were discussed for breaking self-tolerance,
66 leading to autoimmunity causing or supporting the disease pathogenesis (3). Among the
67 autoantibodies, there are “classic” ones which induce immune responses, resulting in the
68 destruction of affected tissues, whereby autoantibodies directed against contractile elements
69 such as anti-myosin and anti-troponin autoantibodies are particularly important (4,5).

70 Starting in the 1970s, the classic autoantibodies were supplemented by an additional class of
71 autoantibodies that bind to G-protein coupled receptors (GPCR-AAB). After receptor binding,
72 GPCR-AAB, in the majority of cases, agonistically activate their related receptors in a similar
73 way to the physiological agonists; therefore GPCR-AAB were called “functional
74 autoantibodies”. However, mechanisms for the prevention of over-boarding receptor
75 stimulation such as receptor down-regulation and desensitization, which are well known with
76 physiological agonists, are lacking for GPCR-AAB. Consequently, GPCR-AAB are discussed
77 as disease drivers as repeatedly summarized in (6-9).

78 With the finding of GPCR-AAB such as directed against the β 1-adrenergic receptor (β 1-AAB)
79 and muscarinic receptor 2 (M2-AAB) (10,11) in patients with DCM of non-ischemic reasons,
80 an autoimmune background of specifically associated with GPCR-AAB emerged. In contrast,
81 patients with ischemic cardiomyopathy and healthy individuals carry GPCR-AAB only in very
82 small amounts or these are completely absent (9).

83 GPCR-AAB directed against the β 1-adrenergic receptor are seen in 70–80% of patients with
84 non-ischemic DCM (6-9).

85 In a distinct group of these patients, preferentially those suffering from arrhythmia, M2-AAB
86 were additionally found with a prevalence of up to 40% (6). Among all heart specific classical
87 and functional autoantibodies, however, the strongest pathogenicity has been demonstrated

88 related to human DCM for β 1-AAB, so that they are increasingly accepted as pathogenic
89 drivers and treatment targets, as summarized in (6-9).

90 Three lines of investigation have proposed the concept of β 1-AAB dependent autoimmunity in
91 the pathogenesis of human DCM; first, experiments using myocardial cells to demonstrate the
92 cardio-pathogenic effects of β 1-AAB at the cellular and subcellular levels; second, animal
93 experiments where rodents were immunized for β 1-AAB generation to demonstrate their
94 cardio-pathogenic effects; and third, and most impressive, clinical trials demonstrating the
95 benefit to DCM patients when specifically their β 1-AAB were removed by immunoadsorption
96 (IA) (12,13). This treatment option is now increasingly accepted for DCM patients positive for
97 β 1-AAB. However, to overcome the problems of IA resulting from costs, logistics and patient
98 burden, drug-associated treatment concepts for the *in vivo* neutralization of β 1-AAB come
99 increasingly to the fore (14,15). Unfortunately, to manifest and extend the knowledge about
100 functional autoantibodies in human DCM in general and specifically of β 1-AAB and even more
101 to prove related treatment concepts in pre-clinical studies, there is a lack of animal models that
102 are more related to human DCM than the rather artificial rodent immunization models (16,17).
103 Although, there are also rodent models with naturally occurring DCM and transgenic mouse
104 lines that were engineered for cardiomyopathy development. There are even mice crossed
105 from transgenic and knockout ones which develop a “so-called” autoimmune cardiomyopathy
106 (18,19). However, there is currently no evidence that such rodents may be suitable to eliminate
107 the lack of models for analyzing the pathogenic role of functional autoantibody associated
108 autoimmunity in human DCM and specifically the role of β 1-AAB and M2-AAB as a driver and
109 treatment target.

110 What's more, “*rodents are phylogenetically very distant from human and some*
111 *pathophysiological features of diseases and their response to pharmacological treatment may*
112 *not be reliable predictors*” (18). Consequently, “*for research aimed at clinical translation, it is*
113 *imperative that initial results from small rodent studies be confirmed in a large animal model*
114 *that more closely resembles humans ...*” (18) and there is “... a simple rule, the closer the
115 heart or body weight of the animal to human heart and body, the more similar are the hearts”

116 (18). Among the large animals that can be used as models for human DCM, Doberman
117 Pinscher (DP) should be of great interest due to their frequent development of dilated
118 cardiomyopathy (DoCM) (20), which has many similarities with human DCM (21-25).
119 DoCM is characterized by three stages. DP in stage one are presumed to have genetic
120 mutations which lead to myocardial alteration on a subcellular level but the majority of cellular
121 changes that occur is still unknown (26,27). However, affected heart mitochondrial protein
122 expression, increased oxidative stress and evidence for apoptosis have been evidenced (28).
123 In this stage, approximately corresponding to NYHA class 1 of human heart failure, the heart
124 is electrically and morphologically normal (23,29). Dogs in state two (occult stage) (NYHA class
125 2) have either ventricular premature complexes (VPC) or a systolic dysfunction, or both, in the
126 absence of overt clinical signs. Dogs in stage three (NYHA class 3/4) present with typical signs
127 very similar as found in human heart failure, such as congestive heart failure (CHF),
128 arrhythmia, syncope and exercise intolerance (20).
129 Here, we demonstrate for the first time that DP frequently carry β 1-AAB that could act as a
130 pathogenic driver in the pathogenesis of cardiomyopathy in a similar way to β 1-AAB in human
131 DCM. Therefore, we suggest that DP could be a suitable model for basic investigation to
132 determine the relationship between β 1-AAB-associated autoimmunity and cardiomyopathy,
133 and even more importantly, to prove treatment concepts to counteract β 1-AAB *in vivo*.

134

135 **Materials and Methods**

136 The study was conducted in accordance with the German animal welfare law. The study
137 protocol was approved by the "Regierung von Oberbayern". DP were enrolled based on owner
138 study agreement.

139

140 **Animals**

141 Client-owned purebred DP attending the Cardiology Department of "Medizinische
142 Kleintierklinik, Ludwig-Maximilians-Universität München" for routine check-up, cardiomyopathy
143 diagnostics or cardiomyopathy follow-up were analyzed for β 1-AAB and M2-AAB.

144 Based on owner study agreement a total of 118 DP (male: n=60; 50.8%, female: n=58; 49.2%)
145 between 1 and 13 years old (median 6 years) were enrolled. To identify DoCM, Holter-ECG
146 was performed for the detection of arrhythmia and echocardiography to evidence cardiac
147 dysfunction. In parallel, blood was sampled for the measurement of functional autoantibodies.
148 Based on the guidelines of the European Society of Veterinary Cardiology (30), DoCM was
149 diagnosed for dogs with >300 VPC/24h or two subsequent examinations within a year showing
150 between 50 and 300 VPC/24h (31) and echocardiographic indicative for cardiac dysfunction.
151 For that purpose, the left ventricular end-systolic (ESVI) and end-diastolic volume (EDVI) were
152 measured and indexed to body surface area based on Simpson's method. An ESVI of >55
153 ml/m² or/and EDVI of >95 ml² were considered to be indicative of DCM.

154

155 ***Measurement of autoantibodies directed against the β 1-adrenergic receptor (β 1-AAB)***
156 ***and muscarinic receptor 2 (M2-AAB)***

157 To measure β -AAB and M2-AAB, a bioassay established by Wallukat and Wollenberger was
158 used (11), which was modified and standardized as described in (32). In this bioassay, the
159 chronotropic response of spontaneously beating cultured neonatal rat cardiomyocytes to the
160 IgG prepared from the dogs' serum was recorded (1 unit of β 1-AAB activity = 1 beat/min
161 frequency change; lower limit of detection (LLD) = 4.0 U; β 1-AAB positivity = \geq 8.0 U). Through
162 the use of specific blockers of the β 1-adrenergic (bisoprolol) and muscarinic receptor 2
163 (atropine), the cells' chronotropic response can be attributed to β 1-AAB or M2-AAB. For
164 comprehensive information about sample (IgG) preparation, bioassay test setup and
165 measurement procedure of GPCR-AAB, see (6,33).

166

167 ***Localization of the receptor binding site with their specific epitope targeted by the***
168 ***autoantibodies directed against the β 1-adrenergic receptor (β 1-AAB) and muscarinic***
169 ***receptor 2 (M2-AAB)***

170 To localize the extracellular binding site (loops), 50 μ l of the autoantibody containing IgG
171 preparation was pre-incubated for 30 min with 2 μ l of solutions containing synthetic peptides

172 (50 $\mu\text{mol/l}$) (Biosyntan GmbH, Berlin-Buch, Germany) which represent the first and second
173 extracellular loops of the β 1-adrenergic and muscarinic receptor 2. Then, this mixture was
174 added to the bioassay for measurement of the autoantibodies' chronotropic activities. To
175 exclusively localize the extracellular binding site of β 1-AAB, the bioassay was performed in the
176 presence of 1 $\mu\text{mol/l}$ atropine to block the M2-AAB activity if present in the IgG preparation. To
177 exclusively localize the target of M2-AAB, the bioassay was performed in the presence of
178 1 $\mu\text{mol/l}$ bisoprolol to block β 1-AAB activity. A comparable procedure was used to map the
179 specific epitope on the receptor loop targeted by the β 1-AAB and M2-AAB. In this case, the
180 bioassay was performed after the pre-treatment of GPCR-AAB containing IgG with an excess
181 of synthetic peptides (Biosyntan GmbH, Berlin-Buch, Germany), which overlapped to
182 represent the amino acid sequence of the receptor loop; first described for β 1-AAB in (34). For
183 mapping of the β 1-AAB targeted epitope on the second extracellular receptor, peptides were
184 used, as follows: P1: HWWRAE, P2: RAESDE, P3: ARRCYND, P4: PKCCDF, and P5:
185 DFVTNR; for M2-AAB epitope mapping: P1: VRTED, P2: EDGECY, P3: CYIQFF, P4: FFSNAA
186 P5: AAVTFG. For this, 50 μl of the GPCR-AAB-containing IgG preparation was pre-incubated
187 for 30 min with 2 μl of solutions containing the synthetic peptides (100 $\mu\text{mol/l}$) (Biosyntan
188 GmbH, Berlin-Buch, Germany). Then, this mixture was added to the Bioassay for GPCR-AAB
189 measurement. In the case of finding the β 1-AAB epitope, the activity was measured as
190 described above in the presence of atropine; for M2-AAB, the bioassay was performed in the
191 presence of bisoprolol.

192

193 ***In vitro indication for the ability to neutralize Doberman pinscher autoantibodies***
194 ***directed against the β 1-adrenergic receptor and muscarinic receptor 2 by drugs***
195 ***already successfully tested in animal and clinical studies for the neutralization of***
196 ***human autoantibodies directed against the β 1-adrenergic receptor***

197 For this purpose, the chronotropic activity of β 1-AAB containing IgG from DP was monitored
198 in the bioassay after pre-incubation of the IgG with drugs that have already been successfully
199 tested in animal and clinical studies for β -AAB inhibition. We tested here a peptide which

200 mimics the amino acid sequence of the second extracellular loop of the β 1-adrenergic receptor
201 (D1) and was synthesized at our request by Biosyntan GmbH, Berlin-Buch, Germany. This
202 peptide acts comparable to the second loop mimics COR-1 which was already studied to
203 counteract β 1-AAB in patients with DCM (14). The other two substances are aptamers (15,35):
204 the first (aptamer 110; D2) is able to neutralize only β 1-AAB due to specific β 1-AAB binding,
205 which was demonstrated *in vitro* and in an animal study (36,37), while the second (BC 007;
206 D3), as demonstrated in animal and human studies, is able to inhibit several GPCR-AAB,
207 including β 1-AAB and M2-AAB (38,39). After drug pre-incubation of β 1-AAB or M2AAB
208 containing IgG from DP with the drugs (test concentration 1 μ mol/l), the mixture was added to
209 the bioassay for measurement of the chronotropic activity of IgG.

210

211 **Statistics**

212 Undetectable marker concentrations (<lower limit of detection, LLD) were numerically
213 expressed as values representing one-half of the LLD. Statistical analysis was performed using
214 the SPSS software package (SPSS Inc., Chicago, US) with Pearson chi-square test and
215 Fisher's exact tests for the comparison of binary variables. For the intergroup comparison of
216 continuous data, the Kruskal-Wallis H-test combined with the Mann-Whitney U-test for post-
217 hoc analysis for the intra-individual comparison of continuous data, and the Friedman test
218 combined with Wilcoxon test for post-hoc analysis was employed.

219 For the graphical representation of continuous patient data, box plots indicate the median and
220 interquartile range (IQR; 25th and 75th percentiles), while whiskers with ends represent the
221 largest and smallest values inside 1.5 times the IQR, outliers (open circles) representing values
222 between 1.5 and 3 times the IQR, and extremes (stars) placed more than 3 times the IQR.

223

224 **Results**

225 **Basic characteristics**

226 Among the study cohort of 118 DP as presented in Table 1, 87 (73.7%) dogs suffered from
227 DoCM which was in age and gender composition comparable to the control group. The

228 cardiomyopathy group consisted of dogs exclusively demonstrating arrhythmias (n=17; 19.5%
229 - VPC/24h: median 205, min 1, max 6465; twice VPC/24 within one year: 286/173/385; EDVI:
230 76.45/55/91, ESVI: 40.51/22/54) indicated as the DoCM-VPC group, with exclusively
231 echocardiographic measures outside of the reference intervals (DoCM-ECHO, n=27; 31.0% -
232 VPC/24h: 5/0/1521; twice VPC/24 within one year: 211/0/1521; EDVI: 107.4/87/196; ESVI:
233 66.24/50/164) as well as those dogs presenting with arrhythmias and echocardiographic
234 pathologies in combination (DoCM-VPC/ECHO, n=43; 49.5% - VPC/24h: 700/0/15 000; twice
235 VPC/24 within one year: 279/124/380; ESVI: 106.8/91/160; EDVI: 67.35/42/106). The groups
236 did not differ significantly in age. In terms of gender composition, the groups DoCM-ECHO,
237 DoCM-VPC/ECHO and the control group are comparable, whereas in the group DoCM-ECHO
238 female animals predominate, especially compared to the group DoCM-ECHO (p>0.05). All
239 dogs were in the pre-clinical, occult stage of the disease. Dogs presenting with severe systemic
240 diseases, end-stage heart failure or non-DCM cardiac diseases were excluded.
241 The group of dogs (n=31; 26.3% of the total number - VPC/24h: 2/0/97; twice VPC/24 within
242 one year: 184/0/279; EDVI: 77.8/53/95; ESVI: 39.3/25/55) which did not fulfill these criteria for
243 DoCM were defined as the primary control group (C). Related to the diagnostic criteria of
244 DoCM used in our study, the control group was composed of healthy animals and those DP at
245 stage 1 of DoCM. At study enrolment, 9 dogs (T0: median age 3 years; min 2, max 3 years)
246 classified into the control group developed cardiomyopathy during the follow-up, diagnosed
247 primarily by VPC detection (T1: median age of 7 years; min 5, max 9 years); they progressed
248 to a diagnosis by the detection of VPCs combined with pathological echocardiography (T2:
249 median age 9 years; min 5, max 9 years).

250

251 ***Autoantibodies directed against the β 1-adrenergic and muscarinic receptor 2 in*** 252 ***Doberman pinschers at the time of enrolment***

253 As indicated in Table 1, 78 (66.1%) of the dogs in the total study cohort presented with β 1-
254 AAB values outside of the reference range ≥ 8 U/min, which means that the dogs were positive
255 for β 1-AAB. Seven dogs also presented with pathological M2-AAB values, and were all also

256 β 1-AAB positive. The rest (n=40; 33.9%) presented with β 1-AAB values in the reference range
257 (< 8U/min). The dogs were sub-divided into those with DoCM and those without signs of DoCM
258 (control group) at enrolment; however, 59 (67.8%) of the dogs with DoCM and 19 (61.3%) of
259 the control group were positive for β 1-AAB. Among the β 1-AAB positive dogs, 5 of the DoCM
260 group and 2 of the control group were also positive for M2-AAB. The remaining 28 (32.2%) in
261 the DoCM group and 12 (38.7%) in the control group were β 1-AAB negative and negative for
262 M2-AAB. Both positivity for β 1-AAB and negativity, respectively, were not significantly different
263 between the groups. However, the median β 1-AAB activity was 19.32 U/min in the DoCM
264 group, which was slightly higher than the 16 U/min reported in the control group.

265 Among the control group, there were 9 dogs, 8 which were positive for β 1-AAB, who developed
266 DoCM in the follow-up. Excluding these dogs from the control group, the statistical evaluation
267 presented a trend (p=0.097) to more β 1-AAB positivity in the DoCM group compared with the
268 control group. When reassembling the DoCM group by supplementing it with the 9 animals
269 developing cardiomyopathy in the follow-up, the tendency (p=0.066) towards more
270 pronounced β 1-AAB positivity in the DoCM group became more marked.

271 Concerning the different DoCM groups, there were no significant differences related to the
272 β 1-AAB positivity (DoCM-VPC: n=12, 70.6%; DoCM-Echo: n=15, 55.6%; DoCM-VPC /Echo:
273 n=32, 74.4%).

274 **Table 1**

275 **Basic characteristics and serum activities of autoantibodies directed against the β 1-**
 276 **adrenergic and muscarinic receptor 2 (* p<0.05; ** p<0.01)**

277

Basic characteristics		Autoantibody presence (n/%)			
		β 1-AAB (n/%)		M2-AAB (n/%)	
		(+)	(-)	(+)	(-)
Total study cohort (n)	118	78/66.1	40/33.9	7/5.9	111/94.1
Age (years; median/min/max)	6/1/13				
Male (n/%)	60/50.8				
Female (n/%)	58/49.2				
Survivor (n/%)	59/50.0	31/52.5	28/47.5	5/8.5	54/91.5
Non-survivor (n/%)	54/45.8	43/79.6**	11/20.4	2/3.7	52/96.3
Lost of follow up (n/%)	5/4.2				
Non-survivor (cardiac reason) (n/%)	35/29.7	26/74.3*	9/25.7	0/0	35/100
Non-survivor (non-cardiac reason) (n/%)	19/16.1	17/89.5*	2/10.5	2/10.5	17/89.5
Doberman cardiomyopathy total (n/%)	87/73.7	59/67.8	28/32.2	5/5.7	83/94.3
Age (years; median/min/max)	7/2/11				
Male (n/%)	46/52.8				
Female (n/%)	41/47.2				
Arrhythmia exclusively (n/%)	17/19.5	12/70.6	5/29.4	1/5.9	16/94.1
Diagnostic criteria					
<i>VPC/24h >300 or Twice 50-300 VPC/24 within one year</i>					
Age (years; median/min/max)	6/2/10				
Male (n/%)	13/76.5				
Female (n/%)	4/23.5				
Medication (n/%)					
<i>No treatment</i>	5/29.4				
<i>Beta-blocker</i>	2/11.8				
<i>Antiarrhythmic drug</i>	2/11.8				
<i>Antiarrhythmic drug + ACE Inhibitor</i>	8/47.0				
Echocardiographic pathologies (n/%)	27/31.0	15/56.6	12/44.4	0/0	27/100
Diagnostic criteria					
<i>ESVI of >55 ml/m² or EDVI of >95 ml²</i>					
Age (years; median/min/max)	8/3/11				
Male (n/%)	8/29.6				

Female (n/%)	19/70.4				
Medication (n/%)					
No treatment	2/7.5				
Calcium sensitizer/PDE 3 inhibitor	12/44.5				
Calcium sensitizer/PDE 3 Inhibitor + ACE Inhibitor	13/48.0				
Arrhythmia + echocardiographic pathologies	43/49.5	32/74.4	11/25.6	4/9.3	39/90.7
Diagnostic criteria					
VPC/24h >300 or Twice 50-300 VPC/24 within one year					
ESVI of >55 ml/m ² or EDVI of >95 ml ²					
Age (years; median/min/max)	7/2/10				
Male (n/%)	20/46.5				
Female (n/%)	23/53.5				
Medication (n/%)					
No treatment	1/2.5				
Calcium sensitizer/PDE 3 inhibitor	1/2.5				
Calcium sensitizer/PDE 3 inhibitor + ACE inhibitor	16/37.2				
Calcium sensitizer/PDE 3 inhibitor + ACE inhibitor + beta-blocker	8/18.6				
Calcium sensitizer/PDE 3 inhibitor + ACE inhibitor + beta-blocker +	13/30.2				
Antiarrhythmic drug	2/4.6				
ACE inhibitor + Antiarrhythmic drug + ACE inhibitor + beta-blocker	2/4.6				
Heathy control group (n/%)	31/26.3	19/61.3	12/38.7	2/6.5	29/93.5
Diagnostic criteria	6/1/13				
Age (years; median/min/max)	17/54.8				
Male (n/%)	14/45.2				
Female (n/%)					

279 ***Follow-up of autoantibodies directed against the β 1-adrenergic receptor in primary***
280 ***healthy dogs who progress to cardiomyopathy***

281 Among the 9 dogs which were healthy at the time of enrolment (DoCM T0) but progressed to
282 DoCM, one of these animals was negative for β 1-AAB at enrolment and remained negative
283 despite progressing to DoCM diagnosed by VPC (DoCM T1) and later than by VPC combined
284 with pathologic echocardiography (DoCM T2). The other 8 dogs were β 1-AAB positive at
285 enrolment. In parallel to the cardiomyopathy development, the β 1-AAB levels increased from
286 T0 to T1 in all DP ($p < 0.05$) and in β 1-AAB positive DP at study enrolment ($p < 0.02$), from T1
287 to T2 in all DP (n.s) and in β 1-AAB positive DP at study enrolment ($p < 0.02$), as well as from
288 T0 to T2 in all DP ($p < 0.05$), and in β 1-AAB positive DP at study enrolment ($p < 0.02$). A further
289 rise in β 1-AAB from T1 to T2 was demonstrated for seven DP. In the one dog that showed a
290 decrease in the β 1-AAB level from T1 to T2, the β 1-AAB level remained clearly in the
291 pathological range (Figure 1).

292 In the comparison of dogs who were free of DoCM for the whole study period with those without
293 the signs of DoCM at enrolment but who progressed to DoCM, a significantly higher proportion
294 of β 1-AAB positivity ($p = 0.044$) was calculated for the last animals.

295

296 **Figure 1**

297 **(A) Activity of autoantibodies directed against the β 1-adrenergic receptor and (B) left**
298 **ventricular end-systolic (ESVI), and end-diastolic volume (EDVI) indexed to body**
299 **surface area and ventricular premature contractions per 24 hours (VPC/24h) in**
300 **primarily healthy Doberman pinschers (DP) (n=9) during the development of severe**
301 **cardiomyopathy (DoCM T0 = healthy; DoCM T1 = cardiomyopathy indicated by arrhythmia;**
302 **DoCM T2 = cardiomyopathy indicated by arrhythmia combined with pathological**
303 **echocardiography); (A) § T1 vs. T0: $p < 0.05$ in all DP, $p < 0.02$ in β 1-AAB positive DP at study**
304 **enrolment; & T2 vs. T1: $p < 0.02$ in β 1-AAB positive DP at study enrolment; # T2 vs. T0:**
305 **$p < 0.05$ in all DP, $p < 0.02$ in β 1-AAB positive DP at study enrolment; (B) § T1 vs. T0: $p < 0.01$**

306 (VPC724h), & T2 vs. T1: $p < 0.02$ (VPC/24h), $p < 0.01$ (EDVI, ESVI), # T2 vs. T0: $p < 0.02$
307 (VPC724h, EDVI), $p < 0.01$ (ESVI).

308

309 ***Mortality of Doberman pinschers related to autoantibodies directed against the β 1-***
310 ***adrenergic receptor***

311 Related to the 118 dogs enrolled, 59 (50%) survived the study period, 35 (29.7%) died due to
312 cardiac reason such as sudden death ($n=30$; 85.7%) or heart failure ($n=5$; 14.3%) and 19
313 (16.1%) died from non-cardiac reasons (Table 1). The median survival time of dogs related to
314 the time of enrolment was 1 year (min 0, max 9 years). Five (4.2%) dogs were lost to follow-
315 up. Of the surviving dogs, 28 (47.5%) were β 1-AAB negative at study enrolment and 31
316 (52.5%) were β 1-AAB positive. In contrast, only 11 (20.4%) of the non-survivors were β 1-AAB
317 negative while 43 (79.6%) were positive for β 1-AAB, which documents a significantly higher
318 prevalence ($p < 0.01$; odd ratio 3.61 (1.57-8.33) of β 1-AAB positivity in the non-survivors.

319 The increased prevalence of β 1-AAB in the non-survivors concerned the dogs that specifically
320 died due to cardiac reasons ($n=9$; 25.7% β 1-AAB negative vs. $n=26$ (74.3%) β 1-AAB positive;
321 $p < 0.05$; odds ratio 2.61 (1.05-6.51) but also those died due to non-cardiac reasons ($p < 0.05$;
322 odds ratio 7.93 (1.68-37.49). With respect to M2-AAB, 54 (91.5%) of the surviving dogs were
323 negative at study enrolment and 5 (5.7%) were positive. However, M2-AAB positivity did
324 significantly increase the risk for death.

325

326 ***Characteristic features of autoantibodies directed against β 1-adrenergic and***
327 ***muscarinic receptor 2 present in Doberman pinschers***

328 As exemplarily demonstrated for β 1-AAB in Figures 2 and 3, both β 1-AAB and M2-AAB found
329 in DP targeted the second extracellular loops of the related receptors. With respect to the
330 epitope on the second extracellular loops which were targeted, the β 1-AAB epitope is located
331 more centrally and contains 3 cysteine residues while the M2-AAB epitope is located closer to
332 the N-terminus and is only flanked by 1 cysteine.

333

334 **Figure 2**

335 **Autoantibodies directed against the β 1-adrenergic receptor (β 1-AAB) of Doberman**
336 **pinschers (n=6) target the second extracellular receptor loop.** Using the bioassay of
337 spontaneously beating cultured neonatal rat cardiomyocytes, the chronotropic activities of
338 the Doberman pinschers' β 1-AAB is demonstrated by the absence or presence of the
339 peptides (L1 = first loop; L2 = second) competing with the first and second extracellular
340 receptor loops. The control experiment was performed in the presence of bisoprolol (BISO).
341 Values below the low limit of detection (LLD) were displayed as half range values. LLD = 4
342 beats/min; cut off (separating healthy from disease subjects) = 8 beats per/min.

343

344 **Figure 3**

345 **Mapping of the second extracellular loop of the β 1-adrenergic receptor for epitope**
346 **localization targeted by the related autoantibodies (β 1-AAB) of Doberman pinschers**
347 Using the bioassay of spontaneously beating cultured neonatal rat cardiomyocytes, the β 1-
348 AAB (n = 5) were measured in the absence or presence of competing peptides that
349 overlapped to represent the second extracellular receptor (P1: HWWRAE, P2: RAESDE, P3:
350 ARRCYND, P4: PKCCDF, and P5: DFVTNR). Values below the low limit of detection (LLD)
351 were displayed as half range values. LLD = - 4 beats/min; cut off (separating healthy from
352 disease subjects) = - 8 beats per/min.

353

354 Based on bioassay measurements, Figure 4 demonstrates that all three drugs which were
355 successful for the neutralization of β 1-AAB in human with DCM were also able to neutralize
356 DP β 1-AAB. Compared with the original β 1-AAB containing DP IgG, the same IgG pre-
357 incubated with the drugs did not present any chronotropic activity on spontaneously beating
358 neonatal cardiomyocytes.

359

360 **Figure 4**

361 **In vitro indication for the ability to neutralize Doberman pinscher autoantibodies**
362 **directed against the β 1-adrenergic receptor by drugs documented to neutralize human**
363 **autoantibodies directed against the β 1-adrenergic receptor.** Using the bioassay of
364 spontaneously beating cultured neonatal rat cardiomyocytes, the β 1-AAB were measured in
365 the absence (control n=8) or presence of the drugs (D1 = second loop peptide (n=4), D2 =
366 aptamer 110 (n=2), D3 = aptamer BC 007 (n=4)). Values below the low limit of detection
367 (LLD) were displayed as half range values. LLD = - 4 beats/min; cut off (separating healthy
368 from disease subjects) = - 8 beats per/min.

369

370 **Discussion**

371 Dilated cardiomyopathy with a disease cumulative prevalence of 58% (20,30,40,41) is the most
372 common form of cardiomyopathy in Doberman pinschers which "... closely resembles the
373 human form of the disease" (24) and therefore repeatedly suggested for modeling human DCM
374 (18,30). In the final state, as mentioned already in above, DP with DoCM present with typical
375 signs such as congestive heart failure (CHF), arrhythmia, syncope and exercise intolerance
376 very similar as found in human heart failure. From anatomical and morphological points of
377 view, left ventricular chamber dilatation and fibrotic cardiac rearrangement were seen (20).
378 The majority of dogs (93.5%) do not survive 2 years after being diagnosed with DoCM (42),
379 which is in agreement with our findings. Despite optimal treatment, the survival of the dogs is
380 about 130 days (median) after entering the overt stage (43).

381 Almost 30 years ago, Smucker et al. (25) suggested that DP should be used as a model for
382 human DCM. Subsequently, DP with dilated cardiomyopathy were announced as "...
383 *remain(ing) (an) untapped resources to investigate both mechanisms of arrhythmias and*
384 *pharmacodynamics of anti-arrhythmics*" (21).

385 However, DP as a DCM model did not gain widespread acceptance in basic research and also
386 not in pre-clinical studies for the testing of human drugs.

387 For the pathogenesis of DoCM, a genetic background is discussed and a autosomal dominant
388 inheritance was proposed (41) but it has been stated (30) that "... *absence of a (specific)*

389 *genetic mutation ... associated with ... DoCM ... does not ensure the dog will never ... develop*
390 *DCM ... (and) ... identification of a genetic mutation does not guarantee the dog will ... develop*
391 *DCM*". Comparable to the genetic background discussed for DoCM, genetic reasons are also
392 assumed to be prominent in human DCM (44). A familial disease history was found in 25% of
393 the human DCM patients (45) detected by a highly diverse genetic background with several
394 gene mutations that, nevertheless, produces a relatively unique DCM phenotype (2).
395 For this complexity in the pathogenesis of DoCM and human DCM, consequently, further
396 causes such as the phenomenon of autoimmunity against the heart must be considered
397 whereby it is currently beyond any doubt that autoimmunity is in tight relation to the individuals'
398 genetic backgrounds. (46). Today, it is increasingly accepted that autoimmunity is as an
399 important pathogenic driver of human DCM (47) and functional autoantibodies such as β 1-
400 AAB and M2-AAB diseases came to the fore (6-9). Finding a comparable autoimmunity in
401 DoCM, additionally to all the other similarities of DoCM with human DCM summarized (21-25),
402 would Doberman pinchers predestinate as model to investigate the autoimmune background
403 of human DCM in general and specifically treatment strategies directed to the functional
404 autoantibodies. We present here for the first time data which indicate that the autoimmune
405 background associated preferentially with β 1-AAB and discussed as driver of human DCM
406 also probably drive the pathogenesis of DoCM.
407 In our study cohort, DP with DoCM showed a prevalence of β 1-AAB of about 70%, comparable
408 as is in human DCM (6). Furthermore, non-surviving dogs were significantly more often positive
409 for β 1-AAB than survivors, which clearly agrees the findings with patients with human DCM
410 studies (48-50).
411 Interestingly, the dogs of our control group also carried β 1-AAB (about 60%). Whether the β -
412 AAB positive dogs are those dogs being genetically compromised for DoCM and therefore in
413 stage one of DoCM remains speculative as it remains whether these dogs are those who
414 progress to the clinically overt DoCM. However, 60 % of β 1-AAB positivity in the control group
415 corresponds to the documented DoCM prevalence (20). Additionally, there was a significantly
416 higher frequency of β 1-AAB positivity in the healthy dogs who developed DoCM during the

417 study period which could support the assumption of β 1-AAB dependent driving to clinically
418 relevant DoCM. Related to humans, we discussed the role of β 1-AAB autoimmunity in
419 progressing to cardiomyopathy for Chagas' patients, where 30% of asymptomatic patients
420 were positive for β 1-AAB and, based on epidemiologic data, nearly 30% of asymptomatic
421 Chagas' patients also progress to Chagas' cardiomyopathy (32). Taking all of this together, we
422 assume a prominent driving role for β 1-AAB associated autoimmunity in the pathogenesis of
423 DoCM, as is increasingly being accepted for human DCM. The resembled role of β 1-AAB
424 autoimmunity in the pathogenesis of DoCM and human DCM, for humans again deduced from
425 Chagas' patient data, was also supported by the increase in the β 1-AAB level from healthy or
426 asymptomatic subjects to those suffering from mild to severe cardiomyopathy, which was seen
427 in both Doberman pinschers and Chagas' patients (32). Consequently, measurement of β 1-
428 AAB could be potentially used for monitoring and prognosis of DoCM and Chagas'
429 cardiomyopathy. Unfortunately, corresponding longitudinal studies focused directly on patients
430 developing DCM are still lacking, but we hope that bio-banking concepts will facilitate the
431 access of such data in the near future.

432 If we take a look at the characteristics of dog and human β 1-AAB, some further analogies were
433 obvious. β 1-AAB of Doberman pinschers target the second extracellular receptor loop, where
434 there is an epitope localized centrally and containing a cysteine between amino acids 193 and
435 204 which is comparable with the epitope targeted by β 1-AAB found in DCM patients (34,51);
436 however, it must be stated that there are additional β 1-AAB in human DCM which target the
437 first extracellular receptor loop (34) which have not been seen in Doberman pinschers.
438 However, β 1-AAB directed against the second extracellular receptor loop were sometimes (52)
439 but not always (34) accused of being the determining cause of DCM.

440 As for β 1-AAB of DCM patients described in (14,37,39), the β 1-AAB activity of Doberman
441 pinschers could be inhibited by peptides which mimic the second extracellular receptor loop or
442 by aptamers binding the autoantibodies. In our view, this is a further indicator that the β 1-AAB
443 associated autoimmunity in Doberman pinschers and human is closely related. Although M2-

444 AAB were found with a clearly lower prevalence in DoCM than published for human DCM, the
445 characteristics are comparable. M2-AAB targeted the second extracellular
446 receptor loop which was also published for M2-AAB of human cardiomyopathies, such as DCM
447 and Chagas' cardiomyopathy (53,54). The specific epitope is located closer to the N-terminus,
448 between amino acids 169 and 177; the same region was also demonstrated for M2-AAB of
449 patients with DCM (unpublished data) or Chagas' cardiomyopathy (54). Related to their
450 specificity for β 1-AAB inhibition, the second loop peptide (D1) and aptamer 110 (D2) did not
451 inhibit M2-AAB, but the aptamer BC 007 (D3), the so-called "broad-band neutralizer" of GPCR-
452 AAB, inhibited the Doberman pinscher M2-AAB as seen for M2-AAB from DCM patients
453 (39,55).

454 Consequently, we suggest, to take the information about functional autoantibody associated
455 autoimmunity in DoCM, together with all the other similarities of DoCM with human DCM as
456 summarized in (21-25), to re-activate Doberman pinschers as a model of human DCM,
457 specifically for basic investigation of the functional autoantibody associated autoimmunity in
458 human DCM and still more importantly for pre-clinical studies in the development of treatment
459 strategies directed against functional autoantibody associated autoimmunity.

460 From our point of view, this is all the more important since, firstly, none of the small animal
461 models used for the modelling of human DCM to date develop functional autoantibodies and,
462 secondly, the functional autoantibodies found in the immunization models seem, based on
463 ELISA experiments (56) to differ from the human autoantibodies in quality and quantity.

464 We agree that a large animal model such as the DP is a priori cost-intensive and there are
465 strong requirements based on "World Medical Statement on Animal Use in Biomedical
466 Research" to respect the welfare of animals in general and specifically of large animals such
467 as DP used for research. However, a study design such as used for the present study with
468 enrolment of client-owned purebred DP attending a veterinary-medical institution for routine
469 check-up, disease diagnostics or follow-up would strongly minimize the cost and guarantee
470 the DP's welfare. In addition, the DP of our study came from different breeding populations
471 throughout Europe and therefore has a greater genetic diversity than the DP from one litter or

472 only a few and can therefore better reflect the pathogenic situation in human DCM with its
473 already mentioned very different genetic background.

474 Last but not least, we have learned that dog owners have a great willingness to participate with
475 their dogs in studies that test new treatment options, especially if it is expected that the
476 treatments can also be beneficial to their dog; always provided that a study design is chosen
477 that guarantees a minimal physiological and psychological impairment of the animals, which
478 was ensured in our study by non-invasive heart examination and only blood analysis.

479

480 **Study limitations**

481 Based on the diagnostic criteria for DoCM used in our study, the control group consisted of
482 healthy animals and DP at stage 1 of DoCM. It is assumed that the dogs at this stage exhibit
483 genetic mutations that lead to myocardial alteration at the subcellular level without becoming
484 electrically or echocardiographically visible. We suspected that the β 1-AAB-positive DP of the
485 control group were those at level 1 of the DoCM. In the future, a detailed characterization of
486 the DP with respect to a genetic predisposition is necessary to verify this speculation.

487

488 **Conclusions**

489 Doberman pinschers with cardiomyopathy presented with typical signs for autoimmunity,
490 preferentially with autoimmunity associated with autoantibodies directed against G-protein
491 coupled receptors, which is closely related to the autoimmunity found in patients with DCM.
492 This, together with the higher prevalence of cardiomyopathy in Doberman pinschers, the fast
493 disease progression and reaching the primary end point of death within around 4 months of
494 entering the severe stage of cardiomyopathy, presents an excellent basis to re-activate
495 Doberman pinschers as a model to study the basics of human DCM. This is specifically the
496 case for GPCR-AAB associated autoimmunity as a disease cause and as a model for pre-
497 clinical studies in drug development aimed at counteracting this form of autoimmunity and
498 consequently DCM.

499

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662

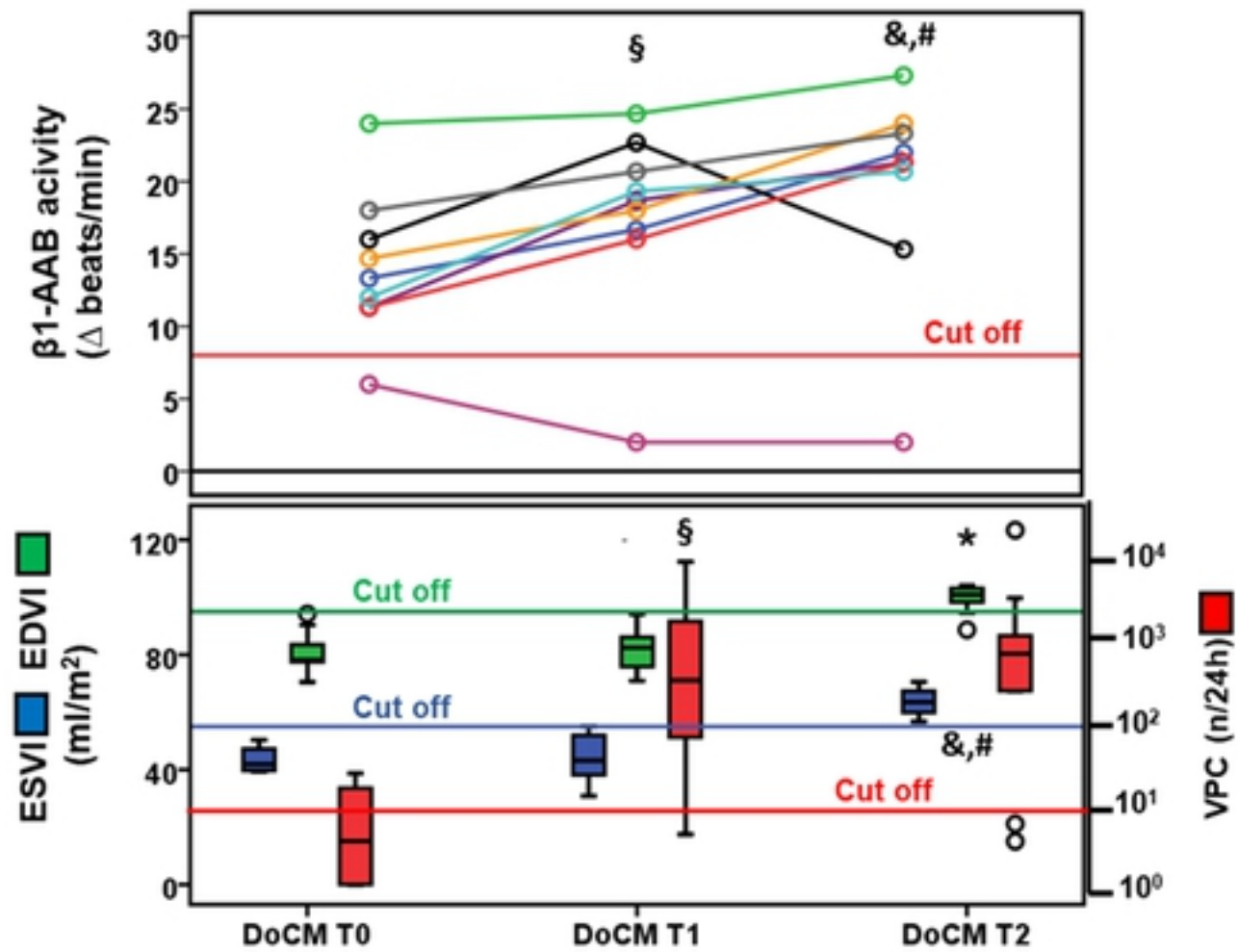


Figure 1

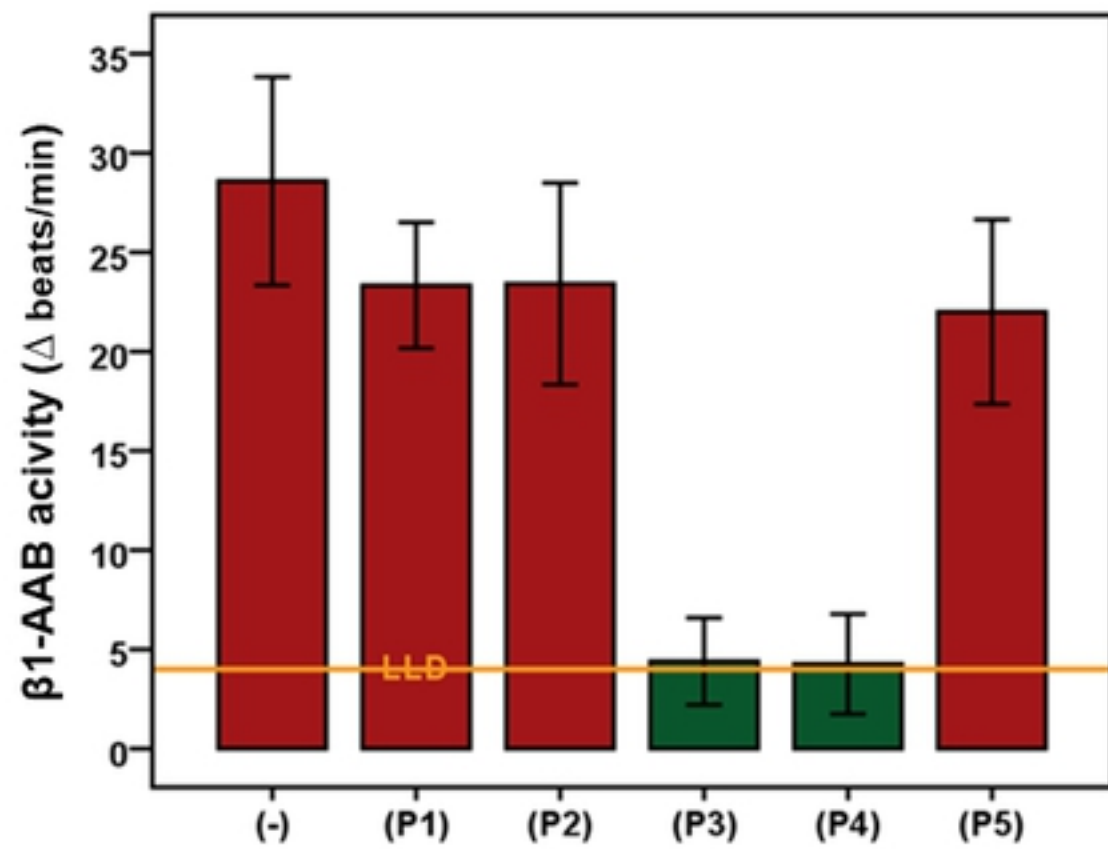


Figure 3

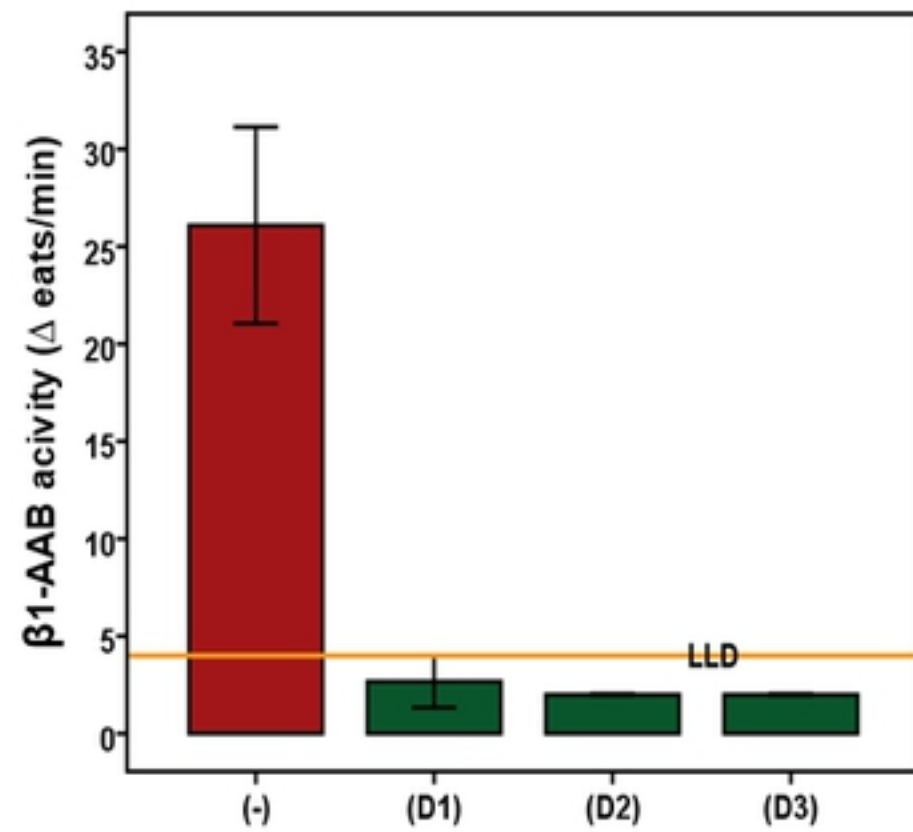


Figure 4

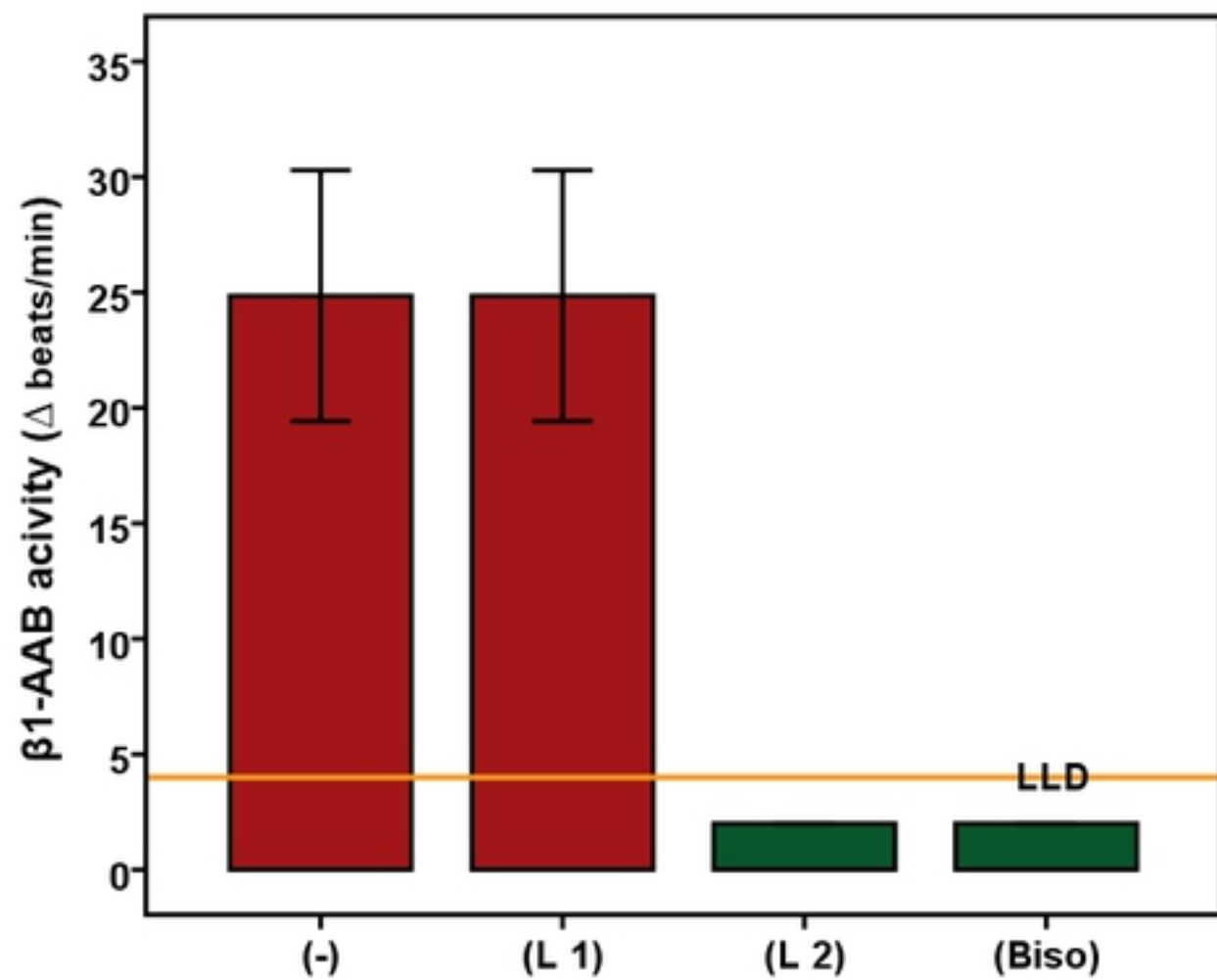


Figure 2